

Feature Articles

Treatment of Relapse of Breast Cancer After Adjuvant Systemic Therapy — Review and Guidelines for Future Research

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INTRODUCTION

Most patients with early breast cancer now receive adjuvant systemic therapy as part of primary treatment. This leads to a significant improvement in survival [1], but there is concern that it may compromise our ability to treat effectively the disease after relapse. The purpose of this review is to examine the available evidence, consider possible mechanisms for an impaired response of advanced disease to treatment and assess the implications for clinical practice.

CLINICAL EXPERIENCE (Table 1)

That adjuvant treatment might be associated with a poor response to treatment on relapse was first suggested in 1981 [2], but, in the same year, others claimed that the response frequency of advanced disease to either endocrine treatment or chemotherapy was not impaired by prior systemic therapy [3]. The first substantial report came from the amalgamated data of two trials of adjuvant cyclophosphamide, methotrexate and 5-fluorouracil (CMF) carried out in Milan [4]. After prior CMF, 38/125 (30%) responded to endocrine treatment compared with 10/25 (40%) who had not had adjuvant treatment. The respective figures for first-line chemotherapy for advanced disease were 32/83 (39%) and 21/55 (38%). The conclusion was that adjuvant chemotherapy did not compromise treatment after recurrence.

Although response frequencies similar to the above were seen at the Mayo Clinic, a different conclusion was reached [5]. In a report of 257 patients who had relapsed after previous adjuvant

treatment with a combination of cyclophosphamide, 5-fluorouracil and prednisolone with or without tamoxifen, the response to endocrine treatment was 47/161 (29%) of median duration 13 months and for chemotherapy 43/156 (28%) of median duration 8 months; median survival from first relapse was 22 months. The response of advanced disease to endocrine treatment was particularly poor if adjuvant treatment had included tamoxifen. Intervening chemotherapy after relapse lowered the response to hormonal therapy, but the converse did not apply.

Clearer evidence of a low response rate to treatment in advanced disease after adjuvant chemotherapy came from experience at Guy's Hospital [6]. Premenopausal patients with stage II breast cancer were randomised to receive postoperative CMF or no adjuvant treatment. 7/38 (18%) patients who relapsed after adjuvant CMF responded to endocrine therapy compared to 23/61 (38%) (P<0.05) who had not had adjuvant treatment. For first-line chemotherapy for advanced disease, the figures were 6/26 (23%) and 24/51 (47%) (P=0.05), respectively. Significant differences in favour of no prior CMF were also found for time to progression (P<0.03), but survival from first relapse was the same irrespective of whether or not adjuvant chemotherapy had been given.

Similar results were reported by the French Epirubicin Study Group [7].477 patients in two consecutive trials received either epirubicin alone or in various combinations with 5-fluorouracil and cyclophosphamide for advanced disease. Stratifying for prior adjuvant CMF, those who received this treatment had a response frequency of 44/137 (32%) compared to 163/340 (48%) (P=0.03) in those without previous adjuvant treatment. In addition to the lower response frequency, prior adjuvant treatment was associated with a significantly shorter time to treatment failure (P=0.002), and reduced survival after relapse (P=0.001). Similar observations were made in another series [8].

Less information is available on the effect of adjuvant endocrine treatment on the responsiveness of advanced disease to systemic therapy. In the Stockholm Breast Cancer Study Group trial, testing adjuvant tamoxifen 40 mg daily for 2 years against no treatment, 378 patients out of 1159 randomised relapsed, of

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Table 1. Results of systemic therapy following recurrence after adjuvant therapy for primary breast cancer

| Reference | Adjuvant therapy | Systemic therapy after relapse | | | | |
|-----------|------------------------|--------------------------------|------------|-----------------------|----------------------|--------------|
| | | Treatment* | n | Responses | Duration (months) | TTP (months) |
| 3 | FAC FAC | CT HT | 38 35 | 15 (39%) 14 (40%) | 16 16 | _ |
| 4 | CMF None | нт нт | 125 25 | 38 (30%) 10 (40%) | 19 17 | _ |
| | CMF None | CT CT | 83 55 | 32 (39%) 21 (38%) | 17 17 16 | _ |
| 5 | CFP ± TAM CFP + TAM | HT CT | 161 156 | 47 (29%) 43 (28%) | - | 13 |
| 6 | CMF None | HT HT | 38 61 | 7 (18%) 23 (38%) | · — | 4 5 |
| | CMF None | CT CT | 26 51 | 6 (23%) 24 (47%) | _ | 2 |
| 7 | CMF None | CT CT | 137 340 | 44 (32%) 163 (48%) | _ | 6 |
| 9 | TAM None | HT HT | 28 26 | 4 (14%) 14 (54%) | _ | 4 15 |
| 10 | TAM TAM | HT CT | 28 44 | 3 (11%) 13 (30%) | _ | _ |

^{*}Only responses to first-line hormonal (HT) or first-line chemotherapy (CT) are given. TTP, median time to progression; FAC, 5 fluorouracil, doxorubicin and cyclophosphamide; CMF, cyclophosphamide, methotrexate and 5-fluorouracil; CFP, cyclophosphamide, 5-fluorouracil and prednisolone; TAM, tamoxifen.

whom 54 were evaluable for the response of advanced disease to either tamoxifen alone or in combination with fluoxymesterone [9]. The objective response rate was significantly lower among patients in the adjuvant tamoxifen group compared to the controls, 4/28 (14%) compared to 14/26 (54%) (P <0.01). The median time to progression was also significantly shorter (4 versus 15 months; P<0.05). In an analysis of 673 patients at Guy's Hospital who received adjuvant tamoxifen, 142 relapsed. Response to endocrine treatment for advanced disease was only 3/28 (11%) and to first-line chemotherapy was 13/44 (30%) [10].

The balance of evidence indicates that adjuvant systemic treatment compromises response of the disease to either endocrine therapy or chemotherapy after relapse. It has also been suggested that patterns of relapses may be affected by prior adjuvant treatment with an increased incidence of liver metastases after chemotherapy, and a higher proportion of patients with pulmonary metastases after endocrine treatment [11]. However, the poorer outcome after relapse does not outweigh the benefits from adjuvant chemotherapy in prolonging survival [1]. On relapse after adjuvant tamoxifen, the disease also appears to be less amenable to endocrine treatment. This applies particularly to the use of tamoxifen, but it is less clear if the response to other agents, such as progestogens or aromatase inhibitors, is also impaired. There is no evidence so far to suggest that responsiveness to chemotherapy is lower after adjuvant endocrine treatment.

MECHANISMS OF IMPAIRED RESPONSE

The biochemical mechanisms of resistance of breast cancer to either chemotherapy or endocrine treatment *in vivo* are not

known. Considerable evidence points to P-glycoprotein over-expression as a mechanism for resistance to various cytotoxic drugs, such as the anthracyclines and vinca alkaloids [12]. Preliminary data suggest that this could explain resistance of breast cancer to these drugs in vivo [13–15], while results from in vitro experiments suggest that other mechanisms could also be involved [16,17]. For endocrine treatment, altered secretion of transforming growth factor- β (TGF- β) by stromal cells [18,19], as well as alterations in intracellular drug metabolism and transport [20], have been suggested as possible mechanisms for tamoxifen resistance. Of particular interest is the lack of cross-resistance between different endocrine treatments, for example, that between tamoxifen and aminoglutethimide [21–23], which is difficult to explain on the basis that both treatments act by depriving tumour cells of oestrogenic stimulation.

In the absence of adequate experimental data, consideration of the mechanisms of impaired response following previous treatment has to be based on theoretical considerations and hypothetical deductions from clinical observations. One possibility is that previous adjuvant therapy could induce acquired resistance and so impede the effect of later treatment. An alternative hypothesis is that a reduced response to later treatment may, at least in part, be due to selection of primary resistant cells. Adjuvant chemotherapy and endocrine therapy each improve long-term and disease-free survival [1]. Whether or not some patients have their micrometastases completely eradicated or tumour growth is delayed for a long time, the numbers of patients who relapse is reduced after adjuvant treatment. Such therapy would, therefore, be expected to reduce both the number and the proportion of patients with tumours sensitive to

treatment among those who relapse (Fig. 1). Thus, the finding of a lower response rate among patients relapsing after adjuvant therapy is not in itself evidence of acquired resistance.

FEASIBILITY OF STUDYING DRUG RESISTANCE

Study of the mechanisms of drug resistance is a major challenge. Primary resistance needs to be evaluated in prospective studies which relate response to systemic treatment and biological parameters measured in biopsies and/or plasma samples obtained immediately before treatment. Any delay between the time of sampling and start of treatment may lead to erroneous conclusions. While oestrogen and progesterone receptors, measured in primary tumours, predict response to endocrine treatment in metastases appearing years later, this may not be so for other parameters. Cancer cells undergo evolutionary changes and adjuvant therapy may alter tumour biology.

As reviewed above, adjuvant therapy influences the response of breast cancer to later treatment, and a high proportion of primary drug-resistant cells in metastases is not likely to be representative of the biology of the untreated primary tumour. There is also the possibility that the metastatic process itself could select cells with particular characteristics such that certain parameters are expressed differently in primary and secondary tumours [24]. Any difference in the expression of biochemical parameters among tumours subsequently responding or not responding to a particular treatment provides no certainty that they represent the mechanism underlying primary resistance. It is well known that expression of certain tumour characteristics such as receptor status, thymidine labelling index, ploidy, genetic alterations, growth factors and histological grading can correlate with each other statistically [25-29], and expression of the mdr phenotype in cells has also been related to biochemical mechanisms of resistance not due to P-glycoprotein overexpression [17,30,31].

Acquired resistance develops over time, and to study it we must assess alterations in tumour biology developing during treatment in individual patients. While primary systemic therapy of primary tumours appears to be an excellent model in which to study primary drug resistance, the treatment of

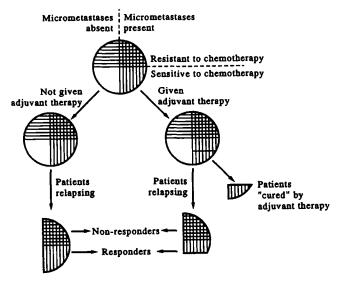


Fig. 1. Illustration of the hypothesis of how adjuvant systemic therapy may lower the proportion of patients responding to later treatment by reducing the incidence of relapse in patients with drug-sensitive tumours.

advanced disease is probably a better one for the study of acquired resistance. For this we need to obtain sequential samples both before starting and during the treatment in order to study biological changes when acquired resistance evolves. In practice, this is difficult to achieve as many patients do not have accessible metastases or it is clinically inappropriate to subject them to repeated biopsies.

Two subgroups of patients with locoregional relapses are suitable for sequential biopsies: patients with multiple, small nodules that may easily be removed, or patients with a single, large tumour suitable for sequential needle biopsies. However, there is always the problem of intratumour variation in the expression of biochemical parameters. While the possibility exists that different nodules may have different biological characteristics, such variation may also exist between different areas within a single tumour mass.

Considering endocrine treatment, many tumours lack oestrogen receptors and are not apparently under endocrine control [31]. These tumours are not expected to respond either to oestrogen deprivation or to anti-oestrogen treatment, but this still occurs in 5–10% of cases. Yet some receptor-positive tumours do not respond to primary endocrine therapy [32–34], and so it is unclear whether or not the mechanisms of primary resistance and acquired resistance in receptor-positive tumours could be similar.

Ideally, studies on the mechanisms of either primary or acquired resistance should be limited to patients treated with single agents given at optimal dosage. This also offers the opportunity to study cross-resistance if a sequential design is used. Biochemical observations in patients treated with combined chemotherapy and hormone therapy would be particularly difficult to interpret. This approach does not conflict with good practice in the management of advanced breast cancer for which the different treatments are generally used alone, sequentially. A particular problem in studying the effects of endocrine treatment is that many patients with advanced disease would have had prior adjuvant tamoxifen, or, if not, tamoxifen is generally given as first-line therapy on relapse. This is likely to impair the study of subsequent forms of endocrine therapy, such as aromatase inhibitors. Apart from possible modifications in tumour biology caused by tamoxifen therapy, a major problem is the long retention time of this drug and its metabolites [35], detectable tissue levels being found in patients several months after terminating treatment [36]. Thus, while it may be important to obtain samples in relation to treatment with aromatase inhibitors or high-dose progestins from patients who have been off tamoxifen for sufficient time, this is not usually practical.

As already mentioned, the use of combination chemotherapy makes it difficult to determine which drug could be responsible for an observed alteration. Some conclusions may be drawn from results obtained in vitro where certain drugs, for example the anthracyclines and vinca alkaloids, have been found to be associated with the mdr phenotype [37], while resistance to other drugs, like the alkylating agents, relates to an increased expression of glutathione sulphotransferases (GSTs) [38]. However, we should be cautious about extrapolating such results to patients. The possibility that a drug may have several mechanisms of action, and that more than one may be responsible for clinical resistance to a drug should always be considered. The anthracycline group of drugs may be used as an example to illustrate this. While these drugs are thought to exert their cytotoxic effect at the DNA level [39–41], other actions have also

been suggested [16,42]. Anthracyclines have been considered to be substrates for the P-glycoprotein pump [43,44], but recent *in vitro* investigations have related doxcrubicin resistance to other mechanisms [45].

An indirect approach could be to compare alterations caused by different drug combinations (like combinations A+B versus combinations A+B+C). However, such comparisons assume the action of the different drugs simply to be additive, and the possibility of pharmacokinetic as well as pharmacodynamic drug interactions should always be kept in mind. However, the use of single agents to treat advanced disease is often justified. Treatment with doxorubicin alone at high dose (70–75 mg/m² every 3 weeks) gives similar response rates to combination chemotherapy, and is suitable for many patients with advanced disease [46,47], providing a more satisfactory opportunity to study the development of resistance which can also be related to dose intensity.

IMPLICATIONS FOR CURRENT PRACTICE

For locoregional relapse after mastectomy and adjuvant treatment, prognostic factors are the disease-free interval, initial clinical stage, histological grade, extent of axillary nodal involvement, the number and location of recurrences, effectiveness and type of local therapy, the use of adjuvant systemic and radiation therapy [48]. Local treatment with surgery or radiation therapy, according to the clinical characteristics of the tumour and previous treatment, is appropriate, but the risk of further recurrence is high, and prognosis is generally poor [48]. Given this poor prognosis after local relapse, it may be reasonable to give adjunctive chemotherapy or hormone treatment to these patients. This approach needs evaluating in clinical trials in which multivariate analyses take note of relevant prognostic factors, including prior adjuvant treatment.

The prognostic significance of local recurrence after mastectomy is worse than after breast conserving treatment; in the latter case, it is often confined to the breast, and there is still the possibility of potentially curative mastectomy [49]. Prognostic factors are the histological type, tumour size at recurrence, diffuse infiltration of the breast or lymphatic involvement. Local treatment is surgical, usually mastectomy, but wide local excision is sometimes possible. As for locoregional relapse after mastectomy, the indications for systemic therapy are ill-defined. Systemic therapy could be considered for poor prognosis patients, preferably within a clinical trial.

Treatment of disseminated disease is medical, and whether to choose chemotherapy of hormone therapy depends on many factors: age and general status of the patient, duration of the relapse-free interval, oestradiol and progesterone receptors, sites of metastatic disease, the presence of clinical symptoms and previous adjuvant cytotoxic or hormone treatment.

For patients relapsing after adjuvant hormonal therapy, the most important clinical parameters to consider are probably the duration of hormonal therapy and disease-free interval. The possibility of further endocrine therapy, for example, with an aromatase inhibitor, should first be considered. When patients have relapsed within 2 years of starting adjuvant endocrine treatment, the responsiveness to another hormonal approach is likely to be poor, although absolute refractoriness is not definite. The clinical decision to use either endocrine therapy or chemotherapy should be based on all available clinical data, particularly the location of metastases and total turnour burden. Extensive visceral involvement is generally an indication for chemotherapy.

After adjuvant chemotherapy, options include either endocrine or further chemotherapy. Important parameters are the disease-free interval, the intensity of prior adjuvant chemotherapy, the drugs used and performance status. In addition to these clinical parameters, oestrogen and progesterone receptors may help in the decision process, but it should be remembered that prior tamoxifen may interfere with the dextran-coated charcoal assay [50,51]. The use of new techniques, utilising monoclonal antibodies recognising receptors irrespective of whether they are bound or unbound to ligand, are to be preferred [52]. It should also be noted that tamoxifen may increase levels of oestrogen and progesterone receptors [53]. If this is a reversible effect, receptor measurements shortly after stopping tamoxifen therapy could give inaccurate predictive information.

For the integration of new cell biological prognostic factors into daily clinical practice, we not only need to know their prognostic power for prediction of relapse-free and overall survival, but also any possible relationship with response to endocrine treatment or chemotherapy, in order for them to be used optimally to select treatment for an individual patient [29,54]. A large number of cell biological parameters are currently available to predict the prognosis of patients with breast cancer, but it is still difficult to predict accurately the response to treatment. A valuable prognostic factor may be a worthless predictive factor to endocrine or chemotherapy, or vice versa. High tumour levels of oestrogen, progesterone and androgen receptors, and PS₂ protein predict a relatively good response to endocrine therapy, while epidermal growth factor receptor positivity, HER2/neu positivity, aneuploidy, high proliferation indices, and possibly high urinary plasminogen activator levels, indicate a likelihood of a poor response to endocrine therapy in metastatic breast cancer. With respect to chemotherapy, a high proliferation rate and HER2/neu amplification predict a good response to therapy in metastatic disease, while mdr gene expression and possibly c-myc amplification are related to a worse response. Thus, we anticipate that the newer cell biological parameters should become increasingly useful in the identification of prognostic subsets of patients and the selection of systemic treatment, and they could also be targets for new approaches to treatment.

Combining endocrine therapy and chemotherapy may yield higher response rates than either used singly [55,56], but in practice such combinations provide additive results at best, and are sometimes partially antagonistic. Thus, endocrine treatments and chemotherapy should preferably be used independently in sequence [57,58]. New promising hormonal or cytotoxic agents could, except in circumstances of clinical urgency, usefully be tested as first-line treatment, standard regimens being used later when needed. This strategy has proved efficient and safe in the development of doxorubicin, epirubicin, mitoxantrone and cisplatin [59–61]. Other clinical studies are needed to define the place of aromatase inhibitors as first-line treatment at the time of relapse in postmenopausal patients, and the role of intensive chemotherapy in selected patients.

CONCLUSIONS

The response of advanced breast cancer to systemic therapy is modified by prior adjuvant treatment. This is, at least in part, a consequence of the selection of primary resistant cells, but the induc ion of acquired resistance may also be important. Whatever the mechanisms, it is important that prior adjuvant treatment should be used as a covariate in the analysis of clinical trials in advanced breast cancer. After adjuvant chemotherapy,

investigational agents should be considered for study earlier rather than later in the clinical course of advanced disease, and the reduced benefits from established drugs should be borne in mind when selecting potentially toxic treatments for palliative use.

- Early Breast Cancer Trialists Collaborative Group. Systemic treatment of early breast cancer by hormonal, cytotoxic, or immune therapy. *Lancet* 1992, 339, 1-15 and 71-85.
- Chlebowski RT, Weiner JM, Luce J, et al. Significance of relapse after adjuvant treatment with combination chemotherapy or 5fluorouracil alone in high risk breast cancer. Cancer Res 1981, 41, 4399-4403.
- 3. Buzdar AU, Legha SS, Hortobagyi GN, et al. Management of breast cancer patients failing adjuvant chemotherapy with adriamycin containing regimens. Cancer 1981, 47, 2798–2802.
- Valagussa P, Tancini G, Bonadonna G. Salvage treatment of patients suffering relapse after adjuvant CMF chemotherapy. Cancer 1986, 58, 1411-1417.
- Buckner JC, Ingle JN, Everson LK, et al. Results of salvage hormonal therapy and salvage chemotherapy in women failing adjuvant chemotherapy after mastectomy for breast cancer. Breast Cancer Res Treat 1989, 13, 135-142.
- Houston SJ, Richards MA, Bentley AE, Smith P, Rubens RD. The influence of adjuvant chemotherapy on outcome after relapse for patients with breast cancer. Eur J Cancer 1993, 29A, 1513-1518.
- Bonneterre J, Mercier M. Response to chemotherapy after relapse in patients with or without previous adjuvant chemotherapy for breast cancer. Cancer Treat Rev 1993, 19, 21-30.
- Falkson G, Gelman R, Falkson CI, Glick J, Harris J. Factors predicting for response, time to treatment failure, and survival in women with metastatic breast cancer treated with DAVTH. J Clin Oncol 1991, 9, 2153-2161.
- Fornander T, Rutqvist LE, Glas U. Response to tamoxifen and fluoxymesterone in a group of breast cancer patients with disease recurrence after cessation of adjuvant tamoxifen. Cancer Treat Rep 1987, 71, 685-688.
- Rubens RD. Effects of adjuvant systemic therapy on response to treatment after relapse. Cancer Treat Rev 1993, 19, 3-10.
- Kamby C, Rose C, Ejlertsen B, et al. Adjuvant systemic treatment and the pattern of recurrences in patients with breast cancer. Eur J Cancer Clin Oncol 1988, 21, 439-447.
- Mattern J, Volm M. Prediction of drug resistance in human tumours using immunohistochemical techniques. Anticancer Res 1992, 12, 413-418
- Schneider J, Bak M, Efferth T, et al. P-glycoprotein expression in treated and untreated human breast cancer. Br J Cancer 1989, 60, 815-818
- Ro J, Sahin A, Ro JY, et al. Immunohistochemical analysis of Pglycoprotein expression correlated with chemotherapy resistance in locally advanced breast cancer. Human Path 1990, 21, 787-791.
- Verrelle P, Meissonnier F, Foncki Y, et al. Clinical relevance of immunohistochemical detection of multidrug resistance P-glycoprotein in breast carcinoma. J Natl Cancer Inst 1991, 83, 111-116.
- Bustamante J, Galleano M, Medrano EE, Boveris A. Adriamycin effects on hydroperoxide metabolism and growth of human breast tumour cells. Breast Cancer Res Treat 1990, 17, 145-153.
- Lee SA, Karaszkiewicz JW, Anderson WB. Elevated level of nuclear protein kinase C in multidrug-resistant MCF-7 human breast carcinoma cells. Cancer Res 1992, 52, 3750-3759.
- 18. Thompson AM, Kerr DJ, Steel CM. Transforming growth factorbeta is implicated in the failure of tamoxifen therapy in human breast cancer. Br J Cancer 1991, 63, 609-614.
- Butta A, MacLennan K, Flanders KC, et al. Induction of transforming growth factor B₁ in human breast cancer in vivo following tamoxifen treatment. Cancer Res 1992, 52, 4261-4264.
- Wiebe VJ, Osborne CK, McGuire WL, DeGregorio MW. Identification of estrogenic tamoxifen metabolite(s) in tamoxifen-resistant human breast tumours. 7 Clin Oncol 1992, 10, 990-994.
- human breast tumours. J Clin Oncol 1992, 10, 990-994.

 21. Buzdar AU, Powell KC, Legha SS, Blumenschein GR. Treatment of advanced breast cancer with aminoglutethimide after therapy with tamoxifen. Cancer 1982, 50, 1708-1712.
- 22. Murray RML, Pitt P. Aminoglutethimide in tamoxifen-resistant

- patients: the Melbourne experience. Cancer Res 1982, 42, 3427s-3441s.
- 23. Kvinnsland S, Lonning PE, Dahl O. Treatment of breast carcinoma with aminoglutethimide. *Acta Radiol Oncol* 1984, 23, 421-424.
- Bradley G, Sharma RK, Rajalakshmi S, Ling V. P-glycoprotein expression during tumour progression in the rat liver. Cancer Res 1992, 52, 5154-5161.
- Silvestrini R, Daidone MG, Di Fronzo G. Relationship between proliferative activity and estrogen receptors in breast cancer. Cancer 1979, 44, 665–670.
- Barbi GP, Marroni P, Bruzzi P, et al. Correlation between steroid hormone receptors and prognostic factors in human breast cancer. Oncology 1987, 44, 265-269.
- Furmanski P, Saunders DE, Brooks SC, Rich MA and the Breast Cancer Prognostic Study Clinical and Pathology Associates. The prognostic value of estrogen receptor determinations in patients with primary breast cancer: an update. Cancer 1990, 46, 2794–2798.
- Jakobsen A, Poulsen HS, Madsen EL, Petersen SE, Hansen HS. Ploidy level of human breast carcinoma. Relation to histopathological features and hormone receptor content. *Acta Radiol Oncol* 1984, 23, 103–107.
- Klijn JGM, Beras EMJJ, Botenbal M, Foekens J. Cell biological factors associated with the response of breast cancer to systemic treatment. Cancer Treat Rev 1993, 19, 45-63.
- Yang L-Y, Trujillo JM, Su Y-Z. Further characterisation of two distinct adriamycin-resistant sublines from LoVo human colon carcinoma cells. *Anticancer Res* 1992, 12, 473–480.
- Kamath N, Grabowski D, Ford J, Kerrigan D, Pommier Y, Ganapathi R. Overexpression of P-glycoprotein and alterations in topoisomerase II in p388 mouse leukaemia cells selected in vivo for resistance to mitoxantrone. Biochem Pharmacol 1992, 44, 937-945.
- McGuire WL. Steroid receptors in human breast cancer. Cancer Res 1978, 38, 4289–4291.
- Degenshein GA, Bloom N, Tobin E. The value of progesterone receptor assays in the management of breast cancer. Cancer 1980, 46, 2789-2793.
- 34. Vollenweider-Zerargui L, Barrelet L, Wong Y, Lemarchand-Beraud T, Gomez F. The predictive value of estrogen and progesterone receptors concentrations on the clinical behaviour of breast cancer in women. *Cancer* 1986, 57, 1171-1180.
- Lonning PE, Lien EA, Lundgren S, Kvinnsland S. Clinical pharmacokinetics of endocrine agents used in advanced breast cancer. Clin Pharmacokinet 1992, 22, 327-358.
- Lien EA, Solheim E, Ueland PM. Distribution of tamoxifen and its metabolites in rat and human tissues during steady state treatment. Cancer Res 1991, 51, 4837–4844.
- Gottesman MM, Pastan I. Clinical trials of agents that reverse multidrug-resistance. J Clin Oncol 1989, 7, 409

 411.
- Waxman DJ. Glutathione S-transferase: role in alkylating agent resistance and possible target for modulation chemotherapy—a review. Cancer Res 1990, 50, 6449-6454.
- Pigram WJ, Fuller W, Hamilton LD. Stereochemistry of intercalation: interaction of daunomycin with DNA. Nature (New Biol) 1972, 235, 17-19.
- Meriwether WD, Bachur NR. Inhibition of DNA and RNA metabolism by daunorubicin in L1210 mouse leukaemia. Cancer Res 1972, 32, 1137-1142.
- Tewey KM, Rowe TC, Yang L, Halligan BD, Liu LF. Adriamycininduced DNA damage mediated by mammalian DNA topoisomerase II. Science 1984, 226, 466–468.
- 42. Tritton TR, Yee G. The anticancer agent adriamycin can be actively cytotoxic without entering cells. *Science* 1982, 217, 248–250.
- Inaba M, Kobayashi H, Sakurai Y, Johnson RK. Active efflux of daunorubicin and adriamycin in sensitive and resistant sublines of p388 leukaemia. Cancer Res 1979, 39, 2200-2203.
- Salmon SE, Grogan TM, Miller T, Scheper R, Dalton WS. Prediction of doxorubicin resistance in vitro in myeloma, lymphoma and breast cancer by P glycoprotein staining. J Natl Cancer Inst 1989, 81.696-701.
- Deffie AM, Alam T, Senevirante C, et al. Multifactorial resistance to adriamycin: relationship of DNA repair, glutathione transferase activity, drug efflux, and P-glycoprotein in cloned cell lins of adriamycin-sensitive and -resistant p388 leukaemia. Cancer Res 1989, 48, 3595-3602.
- Gundersen S, Kvinnsland S, Klepp O, et al. Weekly adriamycin versus VAC in advanced breast cancer. A randomised study. Eur J Cancer Clin Oncol 1986, 22, 1431–1434.

- Richards MA, Hopwood P, Ramirez, et al. Doxorubicin in advanced breast cancer: influence of schedule on response, survival and quality of life. Eur J Cancer 1992, 28A, 1023–1028.
- Recht A, Hayes DF. Local recurrence following mastectomy. In Harris JR, Hellman S, Henderson IC, Kinne DW, eds. Breast Disease, 2nd edition. Philadelphia, Lippincott, 1990, 527-541.
- Recht A, Eberlein TJ, Sadowsky NL. Local recurrence following breast conservation. In Harris JR, Hellman S, Henderson IC, Kinne DW, eds. *Breast Diseases*, 2nd edition. Philadelphia, Lippincott, 1990, 541-547.
- Toma S, Leonessa F, Paridaens R. The effects of therapy on oestrogen receptors in breast cancer. J Steroid Biochem 1985, 23, 1105-1109.
- Toma S, Leclercq G, Legros N, et al. Estrogen receptor variations in neoplastic tissue during the course of disease in patients with recurrent breast cancer. Recent Results Cancer Res 1984, 46, 4233-4236.
- Leclercq G, Bojar H, Goussard J, et al. Abbott monoclonal enzyme immunoassay measurement of oestrogen receptors in human breast cancer: a European multicentre study. Cancer Res 1986, 71, 1266-1272.
- Noguchi S, Motomura K, Imaoka S, Koyama H. Up-regulation of estrogen receptor by tamoxifen in human cancer. Cancer 1993, 71, 1266-1272.

- 54. Klijn JGM, Beras EMJJ, Foekens J. Prognostic factors and response to therapy in breast cancer. Cancer Surveys 1993, in press.
- Cavalli F, Beer M, Martz G. Concurrent or sequential use of cytotoxic chemotherapy and hormone treatment in advanced breast cancer: report of the Swiss group for clinical cancer research. Br Med J 1983, 286, 5-8.
- Mouridsen MT, Rose C, Engelsman E. Combined cytotoxic and endocrine therapy in postmenopausal patients with advanced breast cancer. A randomised study of CMF vs CMF plus tamoxifen. Eur J Cancer Clin Oncol 1985, 12, 291-299.
- Toueini EA. Paridaens R, Heuson JC. Utilisation optimale des combinaisons-hormonales pour le traitement du cancer du sein au stade advance. Nouvelles Perspectives therapeutiques? *Louvain Med* 1986, 105, 111-120.
- Paridaens R. Altering cell kinetics with endocrine therapy. In Henderson IC, ed. Adjuvant Therapy of Breast Cancer. Boston, Kluwer Academic Publishers, 1992, 292–313.
- Perez DJ, Harvey VJ, Robinson BA, et al. A randomised comparison of single-agent doxorubicin and epirubicin as first-line cytotoxic therapy in advanced breast cancer. 7 Clin Oncol 1991, 9, 2148–2152.
- Wilson KS, Paterson AHG. First-line mitoxantrone chemotherapy for advanced breast cancer. Cancer Treat Rep 1986, 70, 1021–1022.
- Sledge GW, Loehrer PJ, Roth BJ, et al. Cisplatin as first-line therapy for metastatic breast cancer. J Clin Oncol 1988, 6, 1811–1814.



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Measurement and Valuation of Quality of Life in Economic Appraisal of Cancer Treatment

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In the economic evaluation (EE) of technologies in cancer treatment at least three endpoints are relevant: costs, survival and quality of life (QoL). This article is focused on QoL. EE requires the use of generic and valuation QoL instruments at a disease non-specific level, but the inclusion of cancer-specific instruments may be advisable, particularly for reasons of explanation if changes in dimensions are small or conflicting. Given the pros and cons of the available questionnaires, we advocate the use of the Nottingham Health Profile, the EuroQol and the Rotterdam Symptom Checklist. In our experience the QoL issue in EE linked with cancer trials is associated with practical problems like questionnaire composition, follow-up time, interviewing schedule, patients' compliance and doctors' acceptance. These problems are discussed and some practical guidelines for the design of QoL measurement in cancer trials are given.

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INTRODUCTION

CANCER IS a major cause of illness and death, responsible for almost a quarter of total mortality in Western countries. During the last few decades cancer treatment has shown a rapid evolution. It is now a multidisciplinary treatment strategy incorporating surgery, radiotherapy, chemotherapy and/or immunotherapy. Treatment usually has important side-effects, especially radiotherapy and chemotherapy. For example, radiotherapy causes tiredness, skin injury and emotional discomfort. Chemo-

therapy, often considered even more burdensome, is given over longer periods and its toxicity causes hair loss, nausea and vomiting, fatigue and emotional problems. Consequently, those involved in the care and treatment of cancer patients have wondered whether improvements in survival probabilities outweigh the burden of these severe side-effects in all cases [1]. Not only life years gained, but also the quality of years alive is at issue.

The high incidence and prevalence of cancer make it a major